

Two Naturally Occurring Intergeneric Hybrid Snakes (*Pituophis catenifer sayi* × *Pantherophis vulpinus*; Lampropeltini, Squamata) from the Midwestern United States

JEFFREY B. LeCLERE,¹ ERICA P. HOAGLUND,^{2,3} JIM SCHAROSCH,⁴ CHRISTOPHER E. SMITH,^{3,5} AND TONY GAMBLE^{6,7,8}

¹878 Galtier Street, Saint Paul, Minnesota 55117 USA

²Minnesota Department of Natural Resources, Division of Ecological and Water Resources—Nongame Program, Saint Paul, Minnesota 55106 USA

³Wildlife Research and Consulting Services, LLC, Saint Paul, Minnesota 55127 USA

⁴811 Boulder Drive, Center Point, Iowa 52213 USA

⁵Department of Fisheries, Wildlife and Conservation Biology, University of Minnesota, Saint Paul, Minnesota 55108 USA

⁶Department of Genetics, Cell Biology and Development, University of Minnesota, Minneapolis, Minnesota 55455 USA

⁷Bell Museum of Natural History, University of Minnesota, Saint Paul, Minnesota 55108 USA

ABSTRACT.—Two intergeneric hybrid snakes (*Pituophis catenifer sayi* × *Pantherophis vulpinus*) are described from the midwestern United States; one collected in south central Iowa and the other from southeastern Minnesota. Both specimens are morphologically intermediate between the putative parental species *P. c. sayi* and *P. vulpinus*. Hybrid origin was verified by comparing DNA sequence data from the hybrids to the putative parental species. Both hybrid specimens possessed *P. c. sayi* mitochondrial DNA haplotypes. Examination of the nuclear gene Vimentin (intron 5) showed both specimens were heterozygous at most variable sites confirming their hybrid origin. These snakes represent only the second and third confirmed instances of naturally occurring intergeneric hybridization among squamate reptile species.

Naturally occurring hybrids among squamate species have been reported with some regularity, but are generally considered uncommon (Murphy and Crabtree, 1988; Campbell et al., 1989; Leaché and Cole, 2007; Mebert, 2008; Kearney et al., 2009). Intergeneric hybridization among squamate species is even more rare, with only three cases reported in the literature. Two of these involved snakes. The first, a cross between *Crotalus horridus* and *Sistrurus catenatus*, was based on detailed morphological evidence with the putative hybrid possessing characteristics intermediate to the parental species (Bailey, 1942). The second case involved a putative *Lampropeltis californiae* × *Pituophis catenifer* cross and featured no formal analysis, simply a photo (Hubbs, 2009). Both instances were based solely on morphology with no genetic data to confirm the morphological hypotheses. The third example involved the hybridization between a Marine Iguana (*Amblyrhynchus cristatus*) and Galapagos Land Iguana (*Conolophus subcristatus*) (Rassmann et al., 1997). Hybrid origin was confirmed by analyzing genetic data with both mitochondrial DNA and restriction fragment length polymorphism (RFLP) analysis of nuclear ribosomal DNA. Examining both morphological and genetic data is the most accurate means to identify putative hybrids in a variety of taxa (Murphy and Crabtree, 1988; Delsuc et al., 2007). Morphological data are essential for making an initial hypothesis of hybridization, as F1 hybrids are generally morphologically intermediate between the two parental species (Bailey, 1942; Hubbs, 1955; Dowling and Secor, 1997). Molecular genetic data can identify parentage and are necessary to verify hybrid origin and confirm the initial morphological identifications. Here we use both morphological and molecular data to describe two instances of naturally occurring intergeneric hybridization in squamates.

MATERIALS AND METHODS

We collected putative *Pituophis catenifer sayi* × *Pantherophis vulpinus* hybrids at two different sites in the midwestern United States, one in south central Iowa and the other in southeastern

Minnesota. Both *P. c. sayi* and *P. vulpinus* occur sympatrically at the localities where the putative hybrids were collected (JBL, JS, CES, and EPH, pers. obs.). We (JBL and JS) collected the first specimen on 16 May 2009 at 1815 hours beneath a large rock on a rock strewn grassy hillside in Madison County, Iowa. The Iowa specimen was captured alive and is currently maintained in captivity (JBL). We (CES and EPH) collected the Minnesota specimen on 18 September 2009 at 2000 hours in Wabasha County, Minnesota. The specimen was found recently hit and dead on the road. We took several photographs in situ and then placed the specimen on ice until tissue could be removed and the specimen could be properly preserved and deposited in the Bell Museum of Natural History, University of Minnesota (JFBM).

We extracted DNA from tissues with the Qiagen DNeasy Blood and Tissue kit (Qiagen, Hilden, Germany) in accordance with the manufacturer's instructions. We amplified a fragment of the mitochondrial *ND4* gene with primers ND4-F (5'-TGA CTA CCA AAA GCT CAT GTA GAA GC-3') (Forstner et al., 1995) and Leu-R (5'-CAT TAC TTT TAC TTG GAT TTG CAC CA-3') (Arevalo et al., 1994). We also amplified intron 5 of the nuclear gene Vimentin with primers VimExon5F (5'-AAC AAT GAT GCC CTG CGC CA-3') and VimExon6R (5'-CAA TAT CAA GAG CCA TCT TTA CAT T-3') (Pyron and Burbrink, 2009). PCR products were purified with the use of Exonuclease I and Shrimp Alkaline Phosphatase (Hanke and Wink, 1994) and sequenced in both directions with the use of Big Dye Terminator 3.1 chemistry on an ABI 3730xl at the Biomedical Genomics Center at the University of Minnesota. Sequences were assembled and checked for accuracy with Sequencher 4.8 (Gene Codes Corp., Ann Arbor, MI).

Newly sequenced material, consisting of one *P. c. sayi*, one *P. vulpinus*, and the two putative hybrids, were combined with sequences from GenBank for all analyses (see Appendix 1). Sequences were aligned with the use of ClustalW (Thompson et al., 1994) and, for the *ND4* data, nucleotides were translated into amino acids with the use of MacClade 4.0.8 (Maddison and Maddison, 1992) to confirm alignment and ensure there were no premature stop codons.

We conducted phylogenetic analysis of the *ND4* data with the use of partitioned maximum likelihood, implemented in

⁸Corresponding author. E-mail: gamb1007@umn.edu
DOI: 10.1670/10-260

TABLE 1. Morphometric and meristic measurements. Data summaries (mean and range) are presented for parental species. All measurements are in millimeters and from the right side unless otherwise noted.

	<i>Pituophis catenifer sayi</i> (N = 12)		<i>P. c. sayi</i> × <i>Pantherophis vulpinus</i> , Iowa specimen		<i>P. c. sayi</i> × <i>P. vulpinus</i> , Minnesota specimen		<i>P. vulpinus</i> (N = 12)	
	Mean	Range					Mean	Range
Ventrals	222.8	213–235	209		206		201.4	189–208
Subcaudals	50.9	37–61	51		55		55.2	48–63
Supralabials	8.3	8–9	8		8		8.1	8–9
Infralabials	10.9	9–12	9		8		10.2	8–11
Prefontals	4.1	3–6	2		3		2.1	2–3
Dorsal scales (row 50)	29.5	28–32	28		27		24.6	22–26
Dorsal blotches	52	45–59	48		52		42.8	37–57
Rostral width	5.2	3.0–8.0	7.35		6.44		7.1	3.5–8.5
Rostral height	6.9	3.8–10.1	6.15		6.22		4.7	2.6–6.4
Prefrontal length	4.8	2.9–6.6	7.07		5.04		5.2	2.8–6.8
Snout length	12.1	6.2–17.3	14.73		12.03		10.3	5.4–13.0
Head length	36.9	20.9–50.3	35.64		38.7		31.2	16.5–39.4
Head width	21.8	11.3–35.3	19.79		19.5		18.1	9.3–22.2
Snout–vent length	1,054	410.0–1,605.0	1,203		1,010		894.2	298–1,210

RAxML 7.0.4 (Stamatakis, 2006). Data were partitioned by codon with a fourth partition for the tRNA. All partitions were assigned the GTR + G model of sequence evolution. We assessed nodal support using 100 nonparametric bootstrap replicates (Felsenstein, 1985). We included additional sequences from *P. c. sayi* and *P. vulpinus* specimens along with sequences from several other *Pituophis* and *Pantherophis* species. *Lampropeltis triangulum* was used as an outgroup.

Phylogenetic analysis of the mitochondrial *ND4* gene allowed us to determine the maternal component of the putative hybrids but could not confirm that they were indeed hybrids. We verified the hybrid origin of the two specimens by examining an independent nuclear locus, intron 5 of Vimentin. We considered heterozygosity in the putative hybrids at the majority of segregating sites between “pure” *P. c. sayi* and *P. vulpinus* as strong evidence for a hybrid origin. We determined heterozygosity by examining chromatograms in Sequencher 4.8 (Gene Codes Corp.). Heterozygous sites were identified easily by the possession of two overlapping peaks at a single locus.

We compared external morphology of the presumed hybrids to putative parental species, *P. c. sayi* and *P. vulpinus*. We collected morphometric and meristic data from 12 *P. c. sayi* and 12 *P. vulpinus* as well as the two putative hybrids (*P. c. sayi* × *P. vulpinus*) (Appendix 1). Measurements were made with the use of Mitutoyo calipers, and all measurements were made to the nearest 0.1 mm. Character descriptions follow Burbrink (2001), and a full list of characters can be found in Table 1. We explored morphological differences among *P. c. sayi*, *P. vulpinus*, and the putative hybrids with the use of principal-components analysis (PCA) with JMP 8 (SAS, 2007). We removed the effect of covariation with snout–vent length (SVL) by using the residuals of the linear regressions between SVL and morphometric measurements.

RESULTS

Maximum-likelihood analysis of 800 base pairs of the mitochondrial *ND4* gene resulted in a well-supported tree (Fig. 1) consistent with recently published molecular phylogenies (Burbrink and Lawson, 2007; Pyron and Burbrink, 2009). Both putative hybrids formed a well-supported clade with other *P. c. sayi* specimens.

We confirmed the hybrid origin of the two snakes by sequencing an approximately 600 base-pair fragment of

Vimentin intron 5. We excluded 40 base pairs of low-quality sequence data from the 3' end of the Iowa hybrid specimen to ensure base call accuracy. Examination of Vimentin intron 5 recovered 13 variable sites, 9 of which were fixed synapomorphies for the *P. c. sayi* and *P. vulpinus* specimens we examined

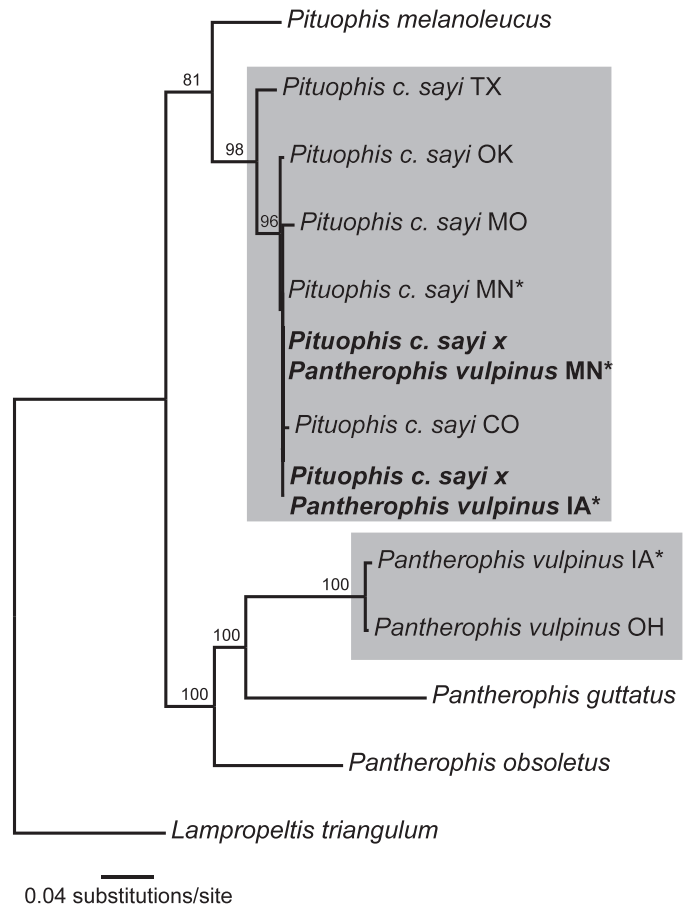


FIG. 1. Phylogenetic relationships among mitochondrial *ND4* haplotypes, constructed with the use of partitioned maximum likelihood (–ln L 2347.270395). *Pituophis catenifer sayi* and *Pantherophis vulpinus* haplotypes are highlighted. Putative hybrid (*P. c. sayi* × *P. vulpinus*) specimens are in bold. Newly sequenced individuals are indicated by an asterisk. Numbers above nodes represent maximum likelihood bootstrap values.

TABLE 2. Variable sites from the Vimentin intron 5 data. Sites representing species-specific synapomorphies are indicated with an asterisk (*).

Sample	30*	42*	69*	149	156*	256*	281*	321	325	408	412*	430*	563*
<i>Pituophis catenifer sayi</i> , Cochise County, Arizona (FJ627902)	G	C	A	T	A	A	T	A	A	A	T	A	C
<i>P. c. sayi</i> Sherburne County, Minnesota (JFBM 16740)	G	C	A	T	A	A	T	T	C	C	T	A	C
<i>Pantherophis vulpinus</i> Ottawa County, Ohio (FJ627910)	A	G	G	G	T	G	A	T	C	A	C	G	G
<i>P. vulpinus</i> Bremer County, Iowa (TG 00132)	A	G	G	T	T	G	A	T	C	A	C	G	G
<i>P. c. sayi</i> × <i>P. vulpinus</i> Madison County, Iowa	A/G	G/C	A/G	T	A/T	A/G	A/T	A/T	A/C	A	C/T	A/G	-
<i>P. c. sayi</i> × <i>P. vulpinus</i> Wabasha County, Minnesota (JFBM 16865)	A/G	C	A/G	T	A/T	A/G	A	A/T	A/C	A	C/T	A/G	G/C

(Table 2). The putative hybrids were heterozygous at most variable sites, e.g., the Minnesota specimen was heterozygous at 69% of variable sites and the Iowa specimen was heterozygous at 83% of variable sites, confirming their hybrid origins.

The two hybrid specimens were morphologically intermediate between *P. c. sayi* and *P. vulpinus*, which was confirmed by the PCA (Figs. 2–4). The first principal component accounted for 41.4% of between-group variability, and the second principal component accounted for 16.1%. Principal component 1 was most strongly loaded among rostral height and rostral width, the number of ventral and prefrontal scales, and the number of dorsal scales. Principal component 2 was most strongly loaded by head length and width and snout length (Table 3).

DISCUSSION

We used both external morphology and molecular genetic data to identify two intergeneric hybrid snakes, *P. c. sayi* × *P. vulpinus*. Both hybrid specimens were intermediate between the parental species in overall appearance. This was particularly evident in cranial morphology, where *P. c. sayi* and *P. vulpinus* are quite different from each other (Fig. 3). Other characteristics were intermediate between the normal ranges for the parental species. The hissing capabilities of the Iowa hybrid, for example,

were midway between the two parental species: longer in duration than the short, forced hiss of *P. vulpinus*, but not nearly as prolonged or raspy as *P. c. sayi* (JBL, pers. obs.). The number of ventral scales and dorsal scale rows in the hybrids were intermediate between the scale counts of the parental species. Not all traits were intermediate, though, and several traits were more closely aligned with one or the other of the parental species. The two hybrids, for example, had divided anal plates as in *P. vulpinus*, compared to the single anal plate of *P. c. sayi*.

The occurrence of *P. c. sayi* × *P. vulpinus* hybrids from two different sites is evidence that isolating mechanisms between *P. c. sayi* and *P. vulpinus* occasionally break down. There are two general categories of isolating mechanisms to prevent hybridization among individuals of different species: prezygotic isolating mechanisms that attempt to prevent mating between different species and postzygotic isolating mechanisms that negatively impact hybrid survival and fertility (Dobzhansky, 1951). Although there is not much information on isolating mechanisms in snakes, there is little evidence to suggest an important role for postzygotic isolating mechanisms. Indeed, many snake species will readily hybridize and produce healthy offspring in captivity, and many of these hybrid offspring are fertile. This even includes captive hybrids among different genera: e.g., *Morelia spilota* × *Liasis mackloti* (Banks and Schwaner, 1984); *Lampropeltis californiae* × *Pantherophis guttatus*; *Pituophis catenifer* × *P. guttatus*; *L. triangulum* × *P. guttatus*; *P. catenifer* × *Pantherophis obsoletus*; *P. catenifer* × *P. vulpinus* (JBL, JS, CES, TG, pers. obs.). Prezygotic isolating mechanisms, on the other hand, likely play a more important role in preventing hybridization among different snake species, including courtship rituals, pheromones, body size, and temporal shifts in

TABLE 3. Character loadings (eigenvectors) for the principal-component (PC) analysis of morphometric and meristic data.

Character	PC 1	PC 2
Ventrals	0.37858	-0.2327
Subcaudals	-0.18868	0.07512
Supralabials	0.13925	-0.04199
Infralabials	0.14064	-0.26451
Prefrontals	0.38481	0.03101
Dorsal scales (50)	0.38755	-0.04305
Dorsal blotches	0.28128	-0.12858
Rostral width (residual)	-0.37863	0.14264
Rostral height (residual)	0.34654	0.2546
Prefrontal length (residual)	-0.25039	0.3013
Snout length (residual)	0.13987	0.50462
Head length (residual)	0.17228	0.466
Head width (residual)	0.17066	0.44876

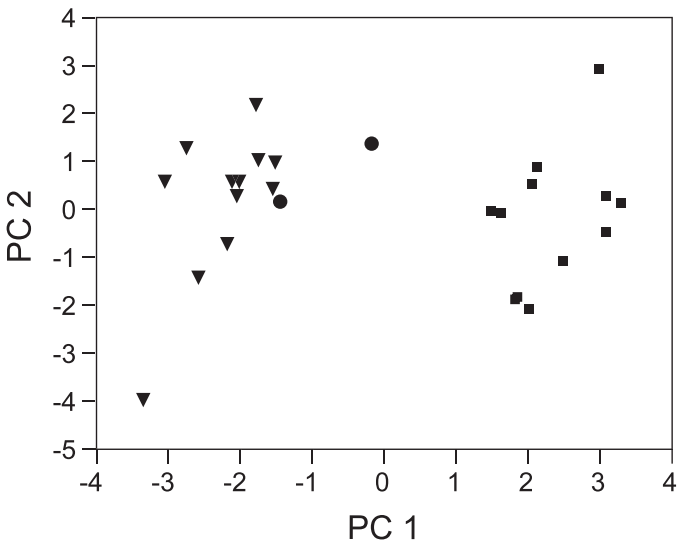


FIG. 2. Principal-component scores for morphological data. Squares represent *Pituophis catenifer sayi*, triangles represent *Pantherophis vulpinus* and circles represent the two *Pituophis c. sayi* × *Pantherophis vulpinus* hybrids.

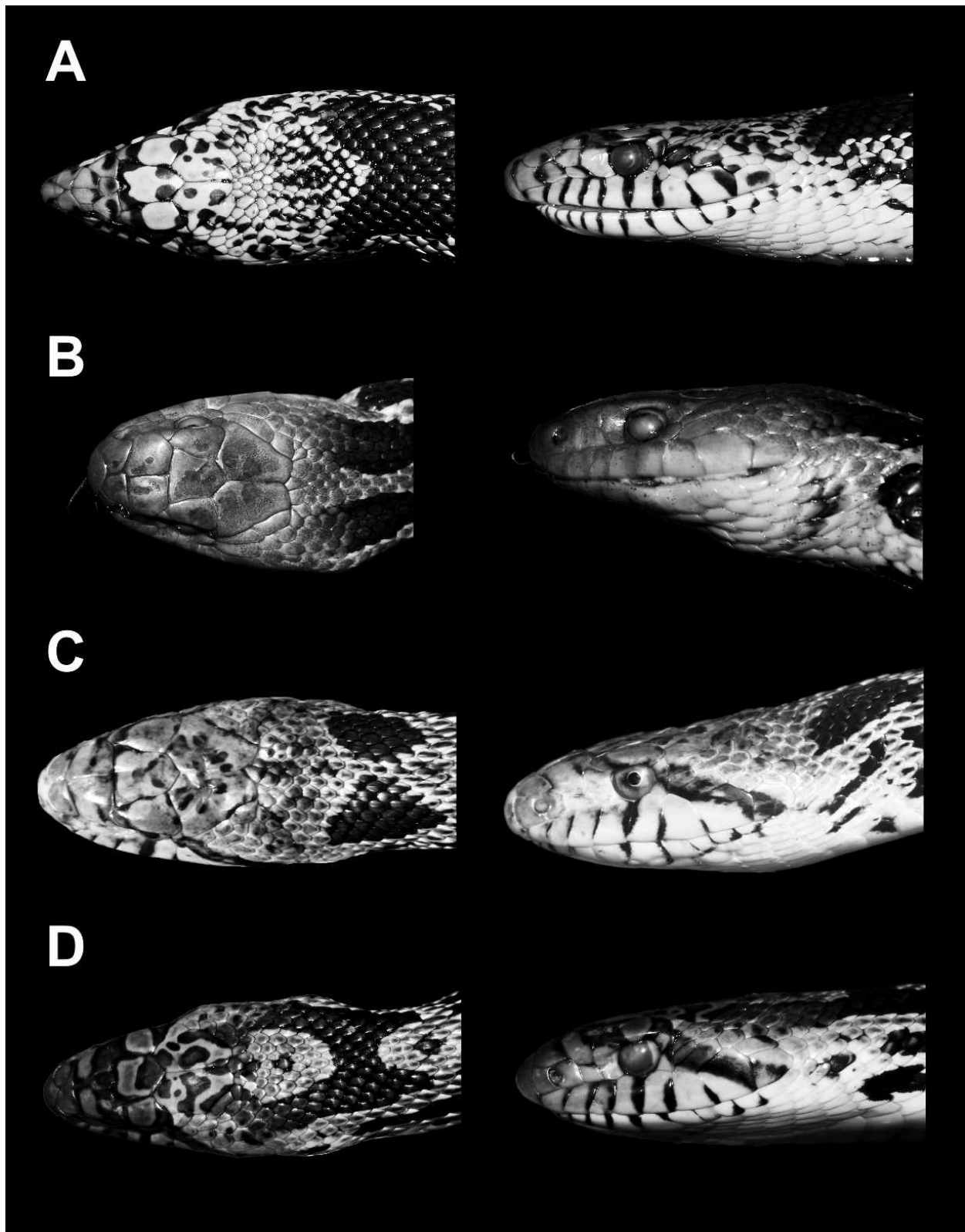


FIG. 3. Dorsal and lateral views of the heads of (A) *Pituophis catenifer sayi*, JFBM 16649, Sioux County, North Dakota; (B) *Pantherophis vulpinus*, JFBM 14521, Renville County, Minnesota; (C) *P. c. sayi* × *Pantherophis vulpinus*, Madison County, Iowa; (D) *Pituophis c. sayi* × *Pantherophis vulpinus* JFBM 16865, Wabasha County, Minnesota. Photos are not to scale.

breeding activity (Shine et al., 2002, 2004). Similarities in mating behavior between *P. c. sayi* and *P. vulpinus* may also explain how such a pairing could occur. Coital biting behavior occurs in both *Pituophis melanoleucus* and *P. vulpinus*, for example, but not in

other studied *Pantherophis* species such as *P. obsoletus* and *P. guttatus* (Shaw, 1951; Gillingham, 1974; Gillingham, 1979; Lewke, 1979). Although we cannot say specifically which of these isolating mechanisms failed, resulting in these two hybrid



FIG. 4. *Pituophis catenifer sayi* × *Pantherophis vulpinus* hybrid from Madison County, Iowa.

individuals, it is interesting to note that both instances involved a male *P. vulpinus* breeding with a female *P. c. sayi*. Male choice appears to be important in preventing hybridization in snakes (Shine et al., 2002, 2004) and these mechanisms apparently broke down on at least two occasions in the midwestern United States.

Hybridization appears to be uncommon among snake species in the wild and is even rarer between snakes in separate genera. Indeed, these are the first instances of intergeneric hybridizations among wild snakes that have been confirmed with genetic data and only the second and third confirmed instances of intergeneric hybridization in a squamate reptile (Rassmann et al., 1997). Despite the broad sympatry and phylogenetic proximity of *P. c. sayi* and *P. vulpinus* (Conant and Collins, 1998; Burbrink and Lawson, 2007), there are no previous reports of natural hybridization between these two species. We are aware of only one successful *P. c. sayi* × *P. vulpinus* breeding in captivity, the result of an accidental pairing in a zoo (M. Edgar, St. Louis Zoo, pers. comm.). Intergeneric hybrids are known among turtles (Karl et al., 1995; Harding and Davis, 1999; Parham et al., 2001); birds (Graves and Zusi, 1990; Grant and Grant, 1992; Graves, 2007); mammals (Jolly et al., 1997; Caballero and Baker, 2010); and are widespread in fish (Hubbs et al., 1988; Burkhead and Williams, 1991; Garrett, 2005). Why intergeneric hybrids appear to be so uncommon in squamates is difficult to determine. This could be partly due to the unequal evolutionary distances between genera in different vertebrate clades (Avisé, 2008). Described squamate genera may simply be older and/or more divergent than teleost, mammalian, or avian genera, making successful intergeneric hybridization among squamates less likely. Given the apparent rarity of intergeneric hybrids in the wild, understanding the exact cause(s) of intergeneric hybridization among squamate species will likely remain elusive for some time.

Acknowledgments.—We thank K. Kozak and A. M. Simons, Bell Museum of Natural History for access to lab space and specimens. K. Mebert and an anonymous reviewer provided useful comments on the manuscript. M. Edgar, St. Louis Zoo for information regarding the zoo's production of *P. c. sayi* × *P. vulpinus* hybrids. M. Pingleton and M. Ricklefs provided help and companionship in the field. The authors complied with the "Guidelines for Use of Live Amphibians and Reptiles in Field

Research" (1987; American Society of Ichthyologists and Herpetologists, the Herpetologists' League, and the Society for the Study of Amphibians and Reptiles) and obtained all applicable state collecting permits.

LITERATURE CITED

- AREVALO, E., S. K. DAVIS, AND J. W. SITES. 1994. Mitochondrial DNA sequence divergence and phylogenetic relationships among eight chromosome races of the *Sceloporus grammicus* complex (Phrynosomatidae) in central Mexico. *Systematic Biology* 43:387–418.
- AVISÉ, J. 2008. Three ambitious (and rather unorthodox) assignments for the field of biodiversity genetics. *Proceedings of the National Academy of Sciences* 105:11564–11570.
- BAILEY, R. M. 1942. An intergeneric hybrid rattlesnake. *American Naturalist* 76:376–385.
- BANKS, C., AND T. D. SCHWANER. 1984. Two cases of interspecific hybridization among captive Australian boid snakes. *Zoo Biology* 3:221–227.
- BURBRINK, F. T. 2001. Systematics of the Eastern Ratsnake complex (*Elaphe obsoleta*). *Herpetological Monographs* 15:1–53.
- BURBRINK, F. T., AND R. LAWSON. 2007. How and when did Old World Ratsnakes disperse into the New World? *Molecular Phylogenetics and Evolution* 43:173–189.
- BURKHEAD, N. M., AND J. D. WILLIAMS. 1991. An intergeneric hybrid of a native minnow, the Golden Shiner, and an exotic minnow, the Rudd. *Transactions of the American Fisheries Society* 120:781–795.
- CABALLERO, S., AND C. S. BAKER. 2010. Captive-born intergeneric hybrid of a Guiana and Bottlenose Dolphin: *Sotalia guianensis* × *Tursiops truncatus*. *Zoo Biology* 29:647–657.
- CAMPBELL, J. A., E. D. BRODIE, D. G. BARKER, AND A. H. PRICE. 1989. An apparent natural hybrid rattlesnake and *Crotalus willardi* (Viperidae) from the Peloncillo mountains of southwestern New Mexico. *Herpetologica* 45:344–349.
- CONANT, R., AND J. T. COLLINS. 1998. *A Field Guide to Reptiles and Amphibians: Eastern/Central North America*. 3rd ed., expanded. Houghton Mifflin, Boston.
- DELSUC, F., M. SUPERINA, G. FERRARIS, M. TILAK, AND E. J. P. DOUZERY. 2007. Molecular evidence for hybridisation between the two living species of South American ratites: potential conservation implications. *Conservation Genetics* 8:503–507.
- DOBZHANSKY, T. 1951. *Genetics and the Origin of Species*. Columbia University Press, New York.
- DOWLING, T. E., AND C. L. SECOR. 1997. The role of hybridization and introgression in the diversification of animals. *Annual Review of Ecology and Systematics* 28:593–619.
- FELSENSTEIN, J. 1985. Confidence-limits on phylogenies—an approach using the bootstrap. *Evolution* 39:783–791.
- FORSTNER, M. R., S. K. DAVIS, AND E. AREVALO. 1995. Support for the hypothesis of anguimorph ancestry for the suborder Serpentes from phylogenetic analysis of mitochondrial DNA sequences. *Molecular Phylogenetics and Evolution* 4:93–102.
- GARRETT, D. L. 2005. A new intergeneric hybrid flatfish (Pleuronectiformes: Pleuronectidae) from Puget Sound and adjacent waters. *Copeia* 2005:673–677.
- GILLINGHAM, J. C. 1974. Reproductive behavior of the Western Fox Snake, *Elaphe v. vulpina* (Baird and Girard). *Herpetologica* 30:309–313.
- GILLINGHAM, J. C. 1979. Reproductive behavior of the rat snakes of Eastern North America, genus *Elaphe*. *Copeia* 1979:319–331.
- GRANT, P. R., AND B. R. GRANT. 1992. Hybridization of bird species. *Science* 256:193–197.
- GRAVES, G. R. 2007. Diagnoses of hybrid hummingbirds (Aves: Trochilidae). 15. A new intergeneric hybrid (*Hylocharis leucotis* × *Selasphorus platycercus*) from the Huachuca Mountains, southeastern Arizona. *Proceedings of the Biological Society of Washington* 120:99–105.
- GRAVES, G. R., AND R. L. ZUSI. 1990. An intergeneric hybrid hummingbird (*Heliodoxa leadbeateri* × *Heliangelus amethysticollis*) from northern Colombia. *The Condor* 92:754–760.
- HANKE, M., AND M. WINK. 1994. Direct DNA-sequencing of PCR-amplified vector inserts following enzymatic degradation of primer and DNTPS. *BioTechniques* 17:858–860.
- HARDING, J. H., AND S. K. DAVIS. 1999. *Clemmys insculpta* (Wood Turtle) and *Emydoidea blandingii* (Blanding's Turtle). Hybridization. *Herpetological Review* 30:225–226.

- HUBBS, B. 2009. Common Kingsnakes: A Natural History of *Lampropeltis getula*. Tricolor Books, Tempe, AZ.
- HUBBS, C., F. B. CROSS, AND F. STEVENS. 1988. Occurrence of natural hybrids between *Etheostoma* and *Percina* (Pisces: Percidae). The Southwestern Naturalist 33:97–99.
- HUBBS, C. L. 1955. Hybridization between fish species in nature. Systematic Zoology 4:1–20.
- JOLLY, C. J., T. WOOLLEY-BARKER, S. BEYENE, T. R. DISOTELL, AND J. E. PHILLIPS-CONROY. 1997. Intergeneric hybrid baboons. International Journal of Primatology 18:597–627.
- KARL, S. A., B. W. BOWEN, AND J. C. AVISE. 1995. Hybridization among the ancient mariners: characterization of marine turtle hybrids with molecular genetic assays. Journal of Heredity 86:262–268.
- KEARNEY, M., M. K. FUJITA, AND J. RIDENOUR. 2009. Lost sex in the reptiles: constraints and correlations. In I. Schön, K. Martens, and P. Dijk (eds.), Lost Sex: The Evolutionary Biology of Parthenogenesis, pp. 447–474. Springer, Dordrecht.
- LEACHÉ, A. D., AND C. J. COLE. 2007. Hybridization between multiple fence lizard lineages in an ecotone: locally discordant variation in mitochondrial DNA, chromosomes, and morphology. Molecular Ecology 16:1035–1054.
- LEWKE, R. E. 1979. Neck-biting and other aspects of reproductive biology of the Yuma Kingsnake (*Lampropeltis getulus*). Herpetologica 35:154–157.
- MADDISON, W. P., AND D. R. MADDISON. 1992. MacClade, Analysis of Phylogeny and Character Evolution. Sinauer, Sunderland, MA.
- MEBERT, K. 2008. Good species despite massive hybridization: genetic research on the contact zone between the watersnakes *Nerodia sipedon* and *N. fasciata* in the Carolinas, USA. Molecular Ecology 17: 1918–1929.
- MURPHY, R. W., AND C. B. CRABTREE. 1988. Genetic identification of a natural hybrid rattlesnake: *Crotalus scutulatus scutulatus* × *C. viridis viridis*. Herpetologica 44:119–123.
- PARHAM, J. F., W. B. SIMISON, K. H. KOZAK, C. R. FELDMAN, AND H. T. SHI. 2001. New Chinese turtles: endangered or invalid? A reassessment of two species using mitochondrial DNA, allozyme electrophoresis and known-locality specimens. Animal Conservation 4:357–367.
- PYRON, R. A., AND F. T. BURBRINK. 2009. Neogene diversification and taxonomic stability in the snake tribe Lampropeltini (Serpentes: Colubridae). Molecular Phylogenetics and Evolution 52:524–529.
- RASSMANN, K., F. TRILLMICH, AND D. TAUTZ. 1997. Hybridization between the Galapagos Land and Marine Iguana (*Conolophus subcristatus* and *Amblyrhynchus cristatus*) on Plaza Sur. Journal of Zoology 242:729–739.
- SAS. 2007. JMP. SAS Institute Inc., Cary, NC.
- SHAW, C. E. 1951. Male combat in American colubrid snakes with remarks on combat in other colubrid and elapid snakes. Herpetologica 7:149–168.
- SHINE, R., R. N. REED, S. SHETTY, M. LEMASTER, AND R. T. MASON. 2002. Reproductive isolating mechanisms between two sympatric sibling species of sea snakes. Evolution 56:1655–1662.
- SHINE, R., B. PHILLIPS, H. WAYE, M. LEMASTER, AND R. T. MASON. 2004. Species isolating mechanisms in a mating system with male mate choice (garter snakes, *Thamnophis* spp.). Canadian Journal of Zoology 82:1091–1098.
- STAMATAKIS, A. 2006. RAXML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics 22:2688–2690.
- THOMPSON, J. D., D. G. HIGGINS, AND T. J. GIBSON. 1994. CLUSTAL W: Improving the sensitivity of progressive multiple sequence alignments through sequence weighting, position specific gap penalties and weight matrix choice. Nucleic Acids Research 22:4673–4680.

Accepted: 12 May 2011.

APPENDIX 1. SPECIMENS EXAMINED

Pituophis catenifer sayi: Morphology—Chisago County, Minnesota: JFBM 13234; Hennepin County, Minnesota: JFBM R1038, JFBM 14536; Houston County, Minnesota: JFBM R1332; Pierce County, Minnesota: JFBM 13235; Sherburne County, Minnesota: JFBM 16740; Goodhue County, Minnesota: JFBM 12712, JFBM 12865, JFBM 12870; Wabasha County, Minnesota: JFBM 16866, JFBM 12181; Sioux County, North Dakota: JFBM 16649. Genetics—Sherburne County, Minnesota: JFBM 16740 (GenBank ND4 JF750662; Vim JF750666).

Pantherophis vulpinus: Morphology—Olmsted County, Minnesota: JFBM 12868; Ramsey County, Minnesota: JFBM R528, JFBM R992; Renville County, Minnesota: JFBM 14521; Rice County, Minnesota: JFBM R367, JFBM R1138; Houston County, Minnesota: JFBM 13530; Sibley County, Minnesota: JFBM 14599; Wabasha County, Minnesota: JFBM R559, JFBM R1170, JFBM 12936, JFBM 16366. Genetics—Bremer County, Iowa TG00132 (GenBank ND4 JF750661, Vim JF750665).

Pituophis c. sayi × *P. vulpinus*: Madison County, Iowa: JBL, live (will be deposited to Drake University upon its demise; GenBank ND4 JF750664; Vim JF750664); Wabasha County, Minnesota: JFBM 16865 (GenBank ND4 JF750663; Vim JF750667).

GenBank Material: ND4—*Pituophis c. sayi* (AF141125, AF141123, AF141124, AF141122); *Pituophis melanoleucus* (AF141111); *Pantherophis vulpinus* (DQ902306); *Pantherophis guttatus* (AM236349); *Pantherophis obsoletus* (DQ902296); *Lampropeltis triangulum* (AY739638). Vimentin (intron 5)—*Pituophis c. sayi* (FJ627902); *Pantherophis vulpinus* (FJ627910).